

Molecular Docking and Target-Based Drug Design of Bioactive Compounds from *Withania somnifera* Against Breast Cancer Targets

Shital D. Tiple^{1*}

¹Institute of Pharmaceutical Education and Research, Borgaon (Meghe), Wardha 442001, Maharashtra, India

*Corresponding Author E-mail: shital.tiple26@rediffmail.com

Abstract:

Breast cancer is the most frequently diagnosed malignancy and the second leading cause of cancer mortality among women worldwide, despite advances in screening, surgery, radiotherapy, endocrine therapy and targeted biological agents. PARC) continued to seek new, safer and multitargeted therapeutics, driven by resistance to existing drugs, toxicity, tumor heterogeneity and the high cost of treatment. Medicinal plants are a valuable source of natural products with bioactive scaffolds that can be rationally optimized through modern computational and structure-based design methodologies. *Withania somnifera* (L.) Dunal, widely known as Ashwagandha, is a primary ingredient of the Ayurvedic system of medication and has been shown to be a key source of steroidal lactones known as withanolides and other active ingredients such as withanosides, sitoindosides and alkaloids that possess various pharmacological effects and influence anticancer activity. In cellular and animal models, before clinical studies or in vitro work of withaferin A, withanolide A, withanone (withania withanolides), ashwagandhanolide and other constituents have demonstrated inhibition of proliferation; induction of apoptosis; suppression of metastasis; and sensitization to chemotherapeutics through the modulation of multiple signaling pathways including NF- κ B, STAT3, Notch, PI3K/Akt/mTOR and ER signaling.

Furthermore, recent advances in molecular docking, molecular dynamics (MD) simulations, quantitative structure activity relationship (QSAR) modeling and network pharmacology have facilitated systematic screening of *W. somnifera* bioactives against a multitude of protein targets implicated in breast cancer pathogenesis such as ER α , HER2, EGFR, phosphatidylinositol-3-kinase α (PI3K α), topoisomerase II α (TOP2A), 17 β -hydroxysteroid dehydrogenase-1 (17 β -HSD1), tumor protein p73 as well as Bcl-2 family members and others. A recent work stands out in this context, where ashwagandhanolide and withanolide sulfoxide were found to be multitarget inhibitors of ER α , 17 β -HSD1, TOP2A and p73 in a PLOS ONE study performed by Lamminga et al., which used docking, MD simulation and ADMET prediction. An article in Drug Design, Development and Therapy (2017), performed docking, QSAR, and ADMET analyses on withanolide analogs for breast cancer targets and outlined structural features associated with activity. This review collates and summarizes the recent bioinformatics data for molecular docking and target-based drug discovery using bioactive compounds of *W. somnifera*, against various breast cancer targets. It outlines breast cancer molecular subtypes and therapeutic targets, *W. somnifera* phytochemistry and anticancer pharmacology, big docking, and modeling studies on ER, HER2/EGFR, PI3K/Akt, topoisomerases, apoptotic regulators such as caspase 8/9 (CASP8/CASP9), BCL-2-associated agonist of cell death (BAD), apoptosis-regulating protein PRAP4P and others. It provides a detailed analysis of the alignment between in silico predictions and experimental data, considers ADMET and drug-likeness factors, and emphasizes future strategies to bridge docking with network pharmacology-based approaches, multi-omics frameworks, as well as by utilizing cutting-edge drug-delivery systems to translate *W. somnifera* phytochemicals into clinically relevant breast cancer therapeutics.

Keywords: *Withania somnifera*; Breast cancer; Molecular docking; Withanolides; Target-based drug design

Received: May 20, 2026

Revised: June 30, 2026

Accepted: July 22, 2026

Published: August 11, 2026

DOI: <https://doi.org/10.64474/3107-6688.Vol2.Issue2.5><https://ddmd.nknpub.com/1/issue/archive>

This is an Open Access article distributed under the terms of the Creative Commons Attribution (CC BY NC), which permits unrestricted use, distribution, and reproduction in any medium, as long as the original authors and source are cited. No permission is required from the authors or the publishers. (<https://creativecommons.org/licenses/by-nc/4.0/>)

1. Introduction

1.1 Breast cancer burden and therapeutic challenges

Breast cancer is responsible for around one in eight diagnoses of cancer globally and is the leading cause of cancer death among women, with increasing incidence in many low- and middle-income countries. Molecular profiling also highlights significant heterogeneity of tumors that are routinely classified into intrinsic subtypes including luminal A and B (ER/PR positive, HER2 negative or low), HER2 enriched, and triple-negative breast cancer (TNBC, (absence of expression for ER, PR and HER2). Each subtype shows unique molecular drivers, clinical behavior, and therapeutic responses¹.

Treatments include surgery, radiotherapy, chemotherapy, endocrine therapy (e.g., tamoxifen, aromatase inhibitors), HER2-targeted agents (trastuzumab, pertuzumab, lapatinib), cyclin-dependent kinase 4/6 inhibitors, PARP inhibitors and immunotherapy. Despite these advances in therapy generating tremendous improvements in patient outcomes, many will ultimately develop resistance, suffer serious toxicity or lack a good pathway for effective targeted therapies including TNBC. Therefore, there is an unmet need for new agents that target multiple pathways, are less toxic and may circumvent or delay resistance²⁻³.

1.2 Natural products and *Withania somnifera* as anticancer leads

Natural products have played an important role in oncology drug discovery, with many of the known chemotherapeutics derived from plants, microbes, and marine organisms. Traditional systems of medicine like Ayurveda offer a rational basis for screening plants that have a history of use against tumor or similar diseases. *W. somnifera*, commonly known as Ashwagandha or Indian ginseng, throughout the ages has remained one of the most popular herbs administered in traditional Indian systems of medicine (Ayurveda) and is a Rasayana (rejuvenative) herb considered beneficial for increasing vitality levels, inducing stress resistance and promoting general health⁴⁻⁵. *W. somnifera* could also be used to treat tumors, inflammatory conditions and neurological disorders.

In the last few decades, phytochemical and pharmacological studies have demonstrated that *W. somnifera* is a rich source of active principles with an array of bioactive compounds that include withanolides (steroidal lactones), withanosides (glycosylated withanolides), sitoindosides, as well as alkaloids; many of these compounds possess significant anticancer activity both in vitro and in vivo. The most characterized derivative compound of *Withania*, withaferin A exhibits

widespread anti-tumor activity against breast, prostate, colon, lung and other neoplasms through modification of various signaling nodes. Withanolides have the potential to take aim at multiple cancer hallmarks, making them appealing candidates for investigation using molecular docking and target-based drug design⁶⁻⁷.

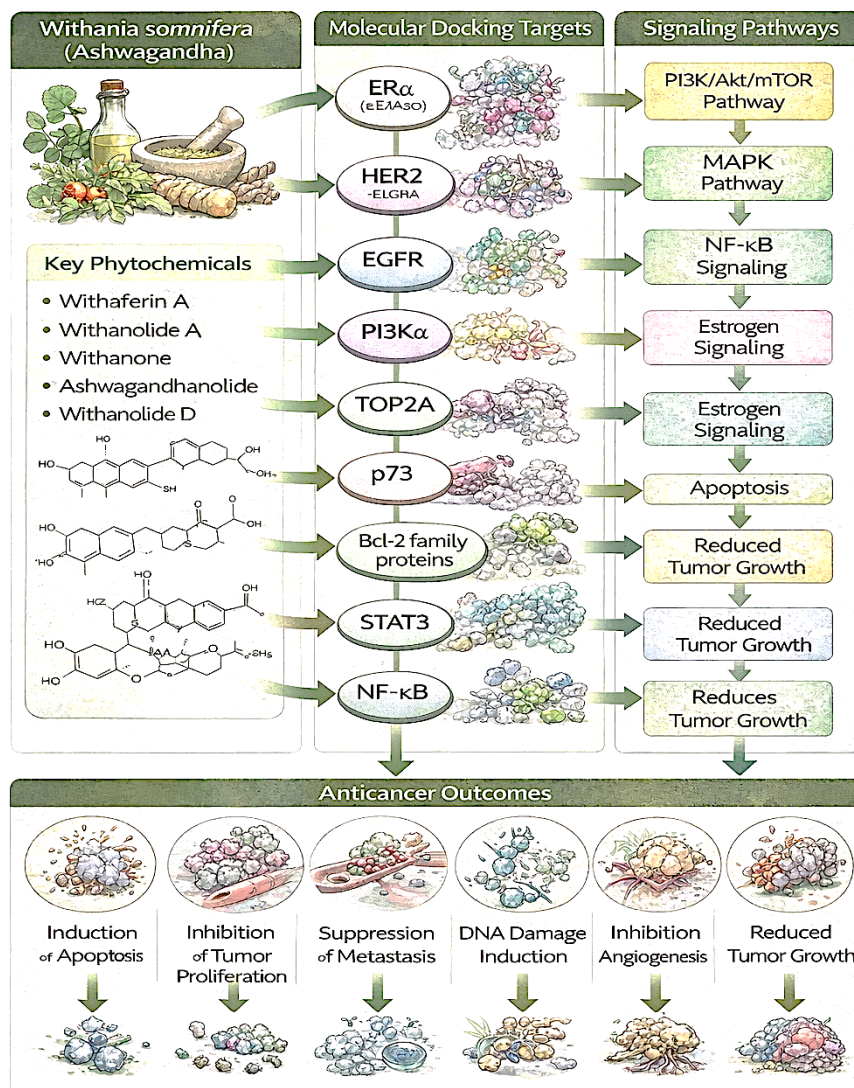


Figure 1. Multi-target anticancer mechanisms of *Withania somnifera* phytochemicals in breast cancer. Major withanolides of *Withania somnifera*, including withaferin A, withanolide A, withanone, and ashwagandhanolide, interact with multiple molecular targets such as ER α , HER2, EGFR, PI3K α , TOP2A, p73, STAT3, and NF- κ B. These interactions regulate key signaling pathways including PI3K/Akt/mTOR, MAPK, NF- κ B, and apoptosis pathways, leading to inhibition of tumor growth, induction of apoptosis, suppression of metastasis, and overall anticancer activity.

Table 1. Major Phytoconstituents of *Withania somnifera* and Their Anticancer Activities in Breast Cancer

Phytochemical Class	Key Compounds	Chemical Nature	Reported Biological Activities	Relevance to Breast Cancer
Withanolides	Withaferin A	Steroidal lactone	Antiproliferative, pro-apoptotic	Inhibits tumor cell growth and induces apoptosis
Withanolides	Withanolide A	Steroidal lactone	Anti-inflammatory, cytotoxic	Suppresses tumor progression
Withanolides	Withanone	Steroidal lactone	ROS generation, apoptosis induction	Selective toxicity toward cancer cells
Withanolides	Ashwagandhanolide	Steroidal lactone	Multitarget inhibition	Interacts with ER α , TOP2A, and p73
Withanolides	Withanolide D	Steroidal lactone	Cytotoxic, anti-metastatic	Inhibits migration and invasion
Glycosides	Withanosides	Glycosylated withanolides	Immunomodulatory	Supports anticancer immune responses
Sitoindosides	Sitoindoside VII–X	Glycosides	Antioxidant, adaptogenic	Protects against oxidative stress
Alkaloids	Anaferine, Cuscohygrine	Nitrogen-containing compounds	Cytotoxic	Enhances anticancer effects

1.3 Role of molecular docking and target-based design

Molecular docking and related structure-based design methods have become central to early-stage drug discovery. Docking mimics small molecule binding onto the active or allosteric sites of target proteins and provides predicted poses and binding affinities that drive hit identification and optimization. For complex herbal mixtures involving herbs such as *W. somnifera*, docking is a powerful approach to provide systematic screening of hundreds of phytoconstituents against multiple breast cancer targets, both identifying promising ligands (small molecules) and informing structure–activity relationships and chemical modifications⁸⁻⁹.

Recent studies have integrated docking with molecular dynamics simulations¹⁰, QSAR (Quantitative Structure–activity relationship), pharmacophore modeling, ADMET prediction

and network pharmacology approaches to generate a more comprehensive picture of *W. somniferous* – breast cancer interactions. These integrative approaches enable rational design of withanolide-derived analogs and formulations that amplify multi-target mechanism(s) while optimizing the pharmacokinetic and safety profiles¹⁰.

2. Phytochemistry and Anticancer Pharmacology of *Withania somnifera*

2.1 Phytochemical profile

W. somnifera, part of the Solanaceae family, contains a variety of withanolides a group of over 300 naturally occurring C-28 steroidal lactones that have an ergostane-based skeleton with a δ -lactone in its side chain. Important withanolides include withaferin A, withanolide A, withanolide D, withanone, 12-deoxywithastramonolide and newer compounds like ashwagandhanolide and withanolide sulfoxide; many of these have been isolated from roots, leaves and berries¹¹.

Withanosides and sitoindosides are glycosidic forms of various withanolides and sterols, whereas minor components comprise alkaloids (anaferine, cuscohygrine), flavonoids and other phenolics. Depending on the plant part, cultivar, geographic origin and extraction method used, the relative abundance of these phytochemicals can vary widely; hence it becomes imperative to standardize pharmacological studies¹².

2.2 General anticancer mechanisms

Multiple reviews describe the anticancer activity of *W. somnifera* and its constituents across various cancer types. Common mechanisms include induction of apoptosis through both intrinsic (mitochondrial) and extrinsic pathways, cell cycle arrest, inhibition of angiogenesis and metastasis, suppression of pro-survival signaling (NF- κ B, STAT3, Akt), generation of reactive oxygen species (ROS), modulation of the heat shock protein (Hsp90) chaperone system, and enhancement of tumor immunogenicity¹³.

In breast cancer, withaferin A has been shown to inhibit NF- κ B activation, downregulate anti-apoptotic proteins (Bcl-2, Bcl-xL, XIAP), upregulate pro-apoptotic factors (Bax, Bak), and activate Notch2/4, leading to reduced proliferation and increased apoptosis in human breast cancer cells. Withanone selectively kills cancer cells by inducing ROS and DNA damage, inhibiting oncogenic transcription factors, and interfering with protein quality control mechanisms. These diverse actions provide multiple entry points for docking-directed target interrogation¹⁴.

2.3 Evidence in breast cancer models

A 2025 in vitro study on MCF-7 breast cancer cells demonstrated that ethanolic *W. somnifera* extract exerts potent cytotoxicity, induces apoptosis via upregulation of p53 and Bax and downregulation of Bcl-2, and enhances the efficacy of doxorubicin in combination regimens, with combination index analysis indicating synergism at specific dose ratios. The extract also elevated caspase-3 activity and increased sub-G1 cell population, supporting apoptosis as the main mode of cell death.¹⁵

Earlier work reports that withaferin A suppresses breast cancer cell proliferation, inhibits migration and invasion by modulating vimentin and filamin, and sensitizes cells to radiation and chemotherapeutics. In xenograft models, withaferin A reduces tumor growth and angiogenesis, partly through inhibition of VEGF-induced endothelial cell functions and disruption of actin cytoskeleton. Together, these findings establish a robust pharmacological foundation for target-based docking investigations¹⁶.

3. Breast Cancer Molecular Targets Relevant to *W. somnifera* Docking

3.1 Hormone receptors: ER α and progesterone receptor

ER α is a nuclear hormone receptor that drives proliferation in approximately 70% of breast cancers. Endocrine therapies such as tamoxifen and aromatase inhibitors target ER signaling but are limited by resistance and side effects. ER α 's ligand-binding domain (LBD) and coactivator binding interface are common docking targets for natural and synthetic ligands¹⁷. 17 β -HSD1, an enzyme that converts estrone (E1) to the more potent estradiol (E2), indirectly enhances ER signaling and is another relevant target for hormone-dependent breast cancer. Docking *W. somnifera* phytochemicals against ER α and 17 β -HSD1 helps identify compounds that may function as selective estrogen receptor modulators (SERMs) or inhibitors of local estrogen biosynthesis¹⁸.

Table 2. Breast Cancer Molecular Targets and Docking Interactions of *Withania somnifera* Phytochemicals

Target Protein	Biological Function	Docked Compounds	Associated Pathway	Therapeutic Significance
ER α (Estrogen receptor alpha)	Hormone receptor signaling	Ashwagandhanolide, Withaferin A	Estrogen signaling pathway	Inhibits hormone-dependent tumor growth
HER2	Receptor tyrosine kinase	Withanolide A	HER2 signaling pathway	Suppresses tumor proliferation
EGFR	Growth factor receptor	Withaferin A	MAPK signaling pathway	Reduces cell proliferation
PI3K α	Cell survival signaling	Withanolide A, Withanone	PI3K/Akt pathway	Inhibits survival pathways

TOP2A	DNA replication enzyme	Ashwagandhanolide	DNA replication pathway	Prevents tumor cell division
p73	Tumor suppressor protein	Withanolide sulfoxide	Apoptosis pathway	Enhances programmed cell death
Bcl-2 family proteins	Apoptosis regulation	Withaferin A	Mitochondrial apoptosis pathway	Promotes apoptosis
STAT3	Transcription factor	Withanolide A	STAT signaling pathway	Suppresses inflammatory signaling
NF-κB	Inflammatory transcription factor	Withaferin A	NF-κB pathway	Reduces tumor inflammation

3.2 Receptor tyrosine kinases: EGFR and HER2

HER2 and EGFR are overexpressed or amplified in subsets of breast cancer and drive aggressive phenotypes via downstream MAPK and PI3K/Akt pathways. Tyrosine kinase inhibitors (TKIs) targeting these receptors have been developed, but resistance and toxicity remain significant issues. Docking *W. somnifera* compounds into the ATP-binding site or allosteric pockets of HER2 and EGFR can identify candidate TKIs or dual inhibitors¹⁹.

3.3 PI3K/Akt/mTOR pathway

PI3K α is frequently mutated or activated in breast cancer, particularly HR-positive tumors, leading to aberrant cell survival and therapy resistance. The catalytic subunit p110 α offers a binding site for ATP-competitive inhibitors, and docking withanolides against PI3K α can reveal potential modulators of this pathway²⁰.

3.4 Topoisomerase II α and DNA repair proteins

TOP2A is essential for DNA replication and chromosome segregation and is a target of chemotherapeutics such as doxorubicin and etoposide. Overexpression of TOP2A is associated with high-risk breast tumors. Docking *W. somnifera* compounds to TOP2A provides insights into potential topoisomerase poisons or catalytic inhibitors²¹.

Other DNA damage response targets include PARP1, BRCA1/2 complexes, and p73, a p53 family member involved in apoptosis and genomic stability. Ashwagandhanolide and related

withanolides have been docked to the tetramerization domain of p73, suggesting possible modulation of its transcriptional activity²².

3.5 Apoptotic and survival regulators

Bcl-2 family proteins, including anti-apoptotic Bcl-2, Bcl-xL, and Mcl-1 and pro-apoptotic Bax and Bak, regulate mitochondrial apoptosis. Withanolides have been proposed to bind Bcl-2-like proteins, shifting the balance toward apoptosis. Heat shock protein Hsp90, NF- κ B transcription factors, and STAT3 are additional survival nodes that have been targeted in docking and modeling studies involving withaferin A and analogs²³.

4. Molecular Docking Studies of *Withania* Phytochemicals Against Breast Cancer Targets

4.1 Docking and MD simulation of ashwagandhanolide and withanolide sulfoxide

A landmark 2022 PLOS ONE study systematically docked 44 phytochemicals from *W. somnifera* against four breast cancer-related proteins: ER α (PDB: 6CBZ), 17 β -HSD1 (1FDW), TOP2A (5GWK), and the p73 tetramerization domain (2WTT). Using AutoDock and other tools, the authors found that ashwagandhanolide and withanolide sulfoxide exhibited the most favorable binding energies and interaction profiles across all four targets²⁴.

Ashwagandhanolide formed stable hydrogen bonds and hydrophobic interactions with key residues in the ER α LBD, overlapping with known ligand-binding sites and suggesting SERM-like potential. Against 17 β -HSD1, it engaged catalytic residues involved in estrone reduction, indicating possible inhibition of local estrogen synthesis. Binding to TOP2A involved intercalation-like positioning in the DNA-binding region, whereas docking to p73's tetramerization domain suggested allosteric modulation²⁵.

MD simulations over 100 ns showed that ashwagandhanolide remained stably bound within the binding cavities of all four proteins, with low RMSD values and favorable binding free energies, especially for ER α and TOP2A complexes. ADMET predictions indicated moderate intestinal absorption, low volume of distribution, ability to interact with ABC transporters, limited BBB penetration, and absence of mutagenicity, hepatotoxicity, or skin sensitization, supporting its drug-likeness for peripheral cancers such as breast cancer²⁶.

4.2 Docking and QSAR of withanolide analogs

A 2017 study by Yadav and collaborators performed docking, QSAR, and ADMET analyses on a panel of withanolide analogs against breast cancer targets, focusing particularly on withaferin A derivatives. Docking indicated strong binding of withaferin A and selected analogs to the ATP-binding pocket of HER2, the hormone-binding pocket of ER α , and the BH3-binding groove of Bcl-xL, with binding energies comparable to or better than reference ligands²⁷.

QSAR modeling correlated structural features such as the presence of specific hydroxyl and epoxy groups, lactone ring configurations, and side-chain modifications with docking scores and known biological activities, identifying pharmacophoric elements critical for target engagement. ADMET predictions suggested reasonable oral bioavailability and acceptable toxicity profiles for optimized analogs, supporting their further development²⁸.

4.3 Systems-level docking and network pharmacology

A 2026 Computational Biology and Medicine article employed a network pharmacology and molecular modeling approach to explore *W. somnifera*-derived phytochemicals against breast cancer at a systems level. The authors compiled bioactive compounds from literature and databases, predicted their targets, and intersected these with breast cancer-associated genes, constructing a compound–target–pathway network²⁹.

Topological analyses identified key targets including ER α , HER2, PI3K, Bcl-B (a Bcl-2 family member), NF- κ B subunits, and cell cycle regulators. Docking and MD simulations confirmed strong and stable binding of selected withanolides, especially withaferin A and withanolide A, to Bcl-B and other hubs, suggesting a role in shifting the apoptosis-survival balance. This network-driven docking strategy helps prioritize high-value targets beyond classic hormone receptors³⁰.

4.4 Docking of withaferin A and withanone to known breast cancer targets

Multiple independent docking studies, summarized in broader reviews, evaluate withaferin A against NF- κ B p65, IKK β , STAT3 SH2 domain, Hsp90, and vimentin. These analyses reveal that withaferin A can covalently or non-covalently bind to cysteine residues in IKK β and vimentin, disrupt Hsp90–client interactions, and block STAT3 dimerization interfaces, thereby inhibiting pro-survival signaling³¹.

In the context of breast cancer, withaferin A has been docked to Notch receptors and γ -secretase components, consistent with experimental observations that it activates Notch2/4-mediated apoptosis in breast cancer cells. Withanone docking studies suggest interactions with DNA-binding domains of oncogenic transcription factors and components of the ubiquitin-proteasome system, supporting its selective toxicity toward cancer cells³².

5. Correlation Between Docking Predictions and Experimental Breast Cancer Data

5.1 ER-related mechanisms

Docking results indicating strong binding of ashwagandhanolide and other withanolides to ER α and 17 β -HSD1 align with experimental data showing anti-estrogenic and antiproliferative effects of *W. somnifera* constituents in hormone-dependent breast cancer cells. For instance, *W. somnifera* extracts reduce ER α expression and estrogen-induced proliferation in MCF-7 cells, and withaferin A downregulates estrogen-responsive genes and synergizes with tamoxifen in vitro³³.

The potential for withanolides to act as SERMs or estrogen biosynthesis inhibitors suggests application in luminal breast cancers, possibly in combination with standard endocrine therapy to mitigate resistance.

5.2 HER2/EGFR and PI3K/Akt pathways

Docking and QSAR work supports the ability of withaferin A and analogs to bind HER2 and PI3K α . Experimental evidence indicates that withaferin A inhibits Akt phosphorylation, reduces mTOR signaling, and decreases HER2 expression in breast cancer models, resulting in decreased proliferation and survival³⁴.

These findings support a multi-target blockade of HER2–PI3K–Akt axes by *W. somnifera* compounds, which could be valuable in HER2-positive and PI3K-mutant tumors, potentially enhancing sensitivity to TKIs or mTOR inhibitors.

5.3 Apoptosis and Bcl-2 family targets

Docking of withanolides to Bcl-B and Bcl-xL in network pharmacology studies is consistent with numerous experiments demonstrating modulation of Bcl-2 family proteins by withaferin A, withanolide A, and withanone. Treatment with these compounds increases Bax/Bcl-2 ratios, triggers mitochondrial membrane depolarization, releases cytochrome c, and activates caspases in breast cancer cells³⁵.

The structural basis for these effects may involve disruption of anti-apoptotic–pro-apoptotic protein interactions or allosteric changes in Bcl-2-like proteins, as suggested by docking models.

5.4 DNA damage and topoisomerase inhibition

Ashwagandhanolide's predicted binding to TOP2A and p73 correlates with observations that withanolides induce DNA damage (e.g., γ -H2AX foci), cause G2/M arrest, and activate p53/p73-driven apoptosis in cancer cells. While direct biochemical evidence of TOP2A inhibition by withanolides is limited, the docking and MD data suggest that ashwagandhanolide may act similarly to topoisomerase poisons or catalytic inhibitors at clinically relevant concentrations³⁶.

6. ADMET, Drug-Likeness, and Scaffold Optimization

6.1 ADMET characteristics of withanolides

Withanolides are relatively lipophilic steroidal lactones, which may aid membrane permeability but raise concerns about solubility and off-target interactions. ADMET analyses in docking studies indicate that many withanolides, including ashwagandhanolide, have moderate intestinal absorption, reasonable oral bioavailability, and limited penetration across the BBB, desirable for breast cancer therapies that do not necessarily require CNS access³⁷.

In silico toxicity predictions generally flag low risks of mutagenicity, cardiotoxicity, and hepatotoxicity, although empirical data remain limited. Metabolic predictions suggest potential interactions with CYP3A4 and other enzymes, underscoring the need for careful drug–drug interaction assessment, particularly when combined with standard chemotherapeutics³⁸.

6.2 Scaffold optimization and analog design

QSAR models from Yadav et al. highlight key structural determinants of activity: the epoxide group at C-5,6, hydroxyl substitutions at specific positions, intact δ -lactone ring, and unsaturation at C-2,3 and C-24,25 correlate with higher docking scores and cytotoxicity. Modifying side chains to fine-tune polarity and reduce metabolic liability has been proposed as a strategy to improve pharmacokinetic properties while preserving target binding³⁹.

Semi-synthetic derivatives that introduce bioisosteric replacements, heterocycles, or conjugation with known pharmacophores (e.g., TKIs) may yield hybrid molecules with enhanced potency or selectivity, guided by docking to multiple targets simultaneously⁴⁰.

7. Methodological Considerations and Limitations of Current Docking Studies

7.1 Protein structures and docking protocols

Many docking studies rely on crystal structures of breast cancer targets complexed with synthetic ligands, such as ER α bound to estradiol or tamoxifen and HER2 bound to TKIs. While these provide high-resolution templates, they may not capture conformational diversity relevant to binding of bulky natural products like withanolides. Ensemble docking using multiple protein conformations and induced-fit protocols could improve accuracy⁴¹.

Details of docking protocols choice of software (AutoDock, Glide, GOLD), grid size, scoring functions, treatment of solvation and protonation vary across studies, complicating direct comparison of binding energies. Validation steps, such as re-docking of co-crystallized ligands and correlation with known IC₅₀ values, are sometimes insufficiently reported⁴².

7.2 Correlation with experimental potency

Docking scores provide relative estimates of binding but do not always correlate linearly with experimental affinity or cellular potency, especially for covalent or allosteric ligands and highly flexible molecules. Withanolides like withaferin A may form covalent adducts with cysteine residues, a mechanism that standard non-covalent docking does not capture accurately. Thus, docking results must be interpreted cautiously and supplemented with MD, free-energy calculations, and experimental binding and functional assays⁴³.

7.3 Complexity of herbal matrices

Most docking studies consider isolated compounds, whereas clinical and traditional usage often involves complex extracts containing multiple phytochemicals. Synergistic or antagonistic interactions between constituents can modulate overall efficacy and toxicity in ways that are not readily predictable from single-compound docking. Systems pharmacology and mixture modeling efforts are needed to capture these emergent properties⁴⁴.

8. Future Directions and Biotechnological Opportunities

8.1 Integrated network pharmacology and multi-omics

Combining docking with network pharmacology, transcriptomics, proteomics, and metabolomics in breast cancer models treated with *W. somnifera* extracts can validate predicted targets, uncover novel mechanisms, and prioritize clinically relevant pathways. Gene expression signatures and pathway activation profiles following withanolide treatment can be mapped onto compound–target networks to distinguish primary from secondary effects⁴⁵.

8.2 Advanced docking and simulation techniques

Future studies should employ ensemble docking, explicit-solvent MD, and free-energy perturbation calculations to refine predictions of ligand–protein binding for withanolides and their analogs. Covalent docking approaches are particularly relevant for electrophilic compounds such as withaferin A, which can form Michael adducts with cysteine residues in IKK β , vimentin, and other proteins⁴⁶.

Machine-learning-based scoring functions and quantum mechanics/molecular mechanics (QM/MM) methods could further improve accuracy when dealing with large, flexible steroidal scaffolds⁴⁷.

8.3 Formulation and targeted delivery

Withanolides are relatively hydrophobic and can be critical for encapsulating into nano-, liposome-, or polymeric micelles. These systems have been proposed for withaferin A and W. somnifera extracts to improve solubility, stability, and tumor accumulation, resulting in enhanced anticancer activity at lower doses. The receptor profile obtained by computational docking of ligands that selectively and specifically target overexpressed receptors (such as HER2, integrins) on the breast cancer cells can be useful in designing targeted delivery systems to achieve localised accumulation of withanolide payloads in tumours⁴⁸.

8.4 Rational combination therapy

The docking and modeling outcomes may help in rational combinations of W. somnifera compounds with standard breast cancer drugs. Docking may reveal non-overlapping binding sites or complementary target spectra between withanolides and agents like doxorubicin, trastuzumab or CDK4/6 inhibitors to inform these studies. Eventually, early in vitro studies already indicated synergistic effects between W. somnifera extract and doxorubicin in MCF-7 cells that could be tailored through target-based design⁴⁹.

8.5 Clinical translation and regulatory aspects

For the potential of W. somnifera phytochemicals to be realized in reducing breast cancer risk or burden, standardized extracts and purified compounds require detailed pharmacokinetic, toxicological and efficacy characterization in appropriate animal models and early-phase human trials. Mechanism-based effects will be demonstrated through biomarker-driven trial designs with molecular stratification (e.g., ER/HER2/PI3K status) and pharmacodynamic readouts (e.g., NF- κ B activity, Bcl-2/Bax ratios).

Regulatory pathways for botanical drugs (for example, the U.S. FDA's Botanical Drug Guidance) demand well-characterized formulations and strong safety and efficacy evidence. Docking and network pharmacology can provide mechanistic insights that boost regulatory dossiers by elucidating target engagement and supporting dose selection⁵⁰.

9. Conclusion

Molecular docking and target-based drug design have led to a significant progress regarding the understanding of interaction between bioactive components of *Withania somnifera* with breast cancer targets. The docking results of withanolides like withaferin A, withanolide A, withanone and ashwagandhanolide showed a correct static binding information into ER α , HER2, EGFR, PI3K α , TOP2A, p73 and other signal proteins like the members of Bcl-2 family that is similar to their antiproliferative pro-apoptotic or chemo sensitizing activities previously found through experiment in breast cancer setting.

Geared towards rational optimization of withanolide scaffolds and the development of novel analogs with improved potency and selectivity, results from docking studies, MD simulations,

QSAR analysis and ADMET predictions underline structural characteristics as well as pharmacokinetic properties that can be exploited for this purpose. Network pharmacology methods further elucidate that phytochemicals from *W. somnifera* alter interwoven pathway controlling hormone signaling, cell survival, apoptosis and DNA repair in support of a poly-targeting therapeutic paradigm aptly suited for the heterogeneous and adaptive characteristics of breast cancer.

However, computational results need to be interpreted with caution considering methodological caveat and inherent complexity of herbal mixtures pharmacology. Comprehensive and empirical validation of putative therapeutic targets; PK/PD & toxicity assessment; as well as carefully designed human clinical studies will be required to convert the promise of *W. somnifera* bioactive compounds into safe and effective therapies for breast cancer. The marriage of advanced in silico modeling with experimental and clinical studies makes it possible to transform the traditional glory of Ashwagandha into modern, evidence-based, multi-target agents for personalized breast cancer care.

References

1. Najib Nik A Rahman, N., Furuta, T., Kojima, S., Takane, K., & Ali Mohd, M. (1999). Antimalarial activity of extracts of Malaysian medicinal plants. *Journal of Ethnopharmacology*, 64, 249–254. [https://doi.org/10.1016/S0378-8741\(98\)00135-4](https://doi.org/10.1016/S0378-8741(98)00135-4)
2. Nguyen, P. H., Tran, V. D., Pham, D. T., Dao, T. N. P., & Dewey, R. S. (2021). Use of and attitudes towards herbal medicine during the COVID-19 pandemic: A cross-sectional study in Vietnam. *European Journal of Integrative Medicine*, 44, 101328. <https://doi.org/10.1016/j.eujim.2021.101328>
3. Nguyen, T. K., Tran, L. T. T., Ho, D. V., Thai, P. H., Ha, T. P., Ty, P. V., et al. (2023). Xanthine oxidase, α -glucosidase and α -amylase inhibitory activities of the essential oil from *Piper lolot*: In vitro and in silico studies. *Heliyon*, 9, e19148. <https://doi.org/10.1016/j.heliyon.2023.e19148>
4. Ojo, O. A., Adegboyega, A. E., Johnson, G. I., Umedum, N. L., Onuh, K., Adeduro, M. N., et al. (2021). Deciphering the interactions of compounds from *Allium sativum* targeted towards identification of novel PTP1B inhibitors in diabetes treatment: A computational approach. *Informatics in Medicine Unlocked*, 26, 100719. <https://doi.org/10.1016/j.imu.2021.100719>
5. Onikanni, A. S., Lawal, B., Oyinloye, B. E., Mostafa-Hedeab, G., Alorabi, M., Cavalu, S., et al. (2022). Therapeutic efficacy of *Clompanus pubescens* leaves fractions via downregulation of neuronal cholinesterases/ $\text{Na}^+/\text{K}^+\text{ATPase}$ /IL-1 β , and improving neurocognitive and antioxidant status of streptozotocin-induced diabetic rats. *Biomedicine & Pharmacotherapy*, 148, 112730. <https://doi.org/10.1016/j.biopha.2022.112730>
6. Othman, N. S., Che Roos, N. A., Aminuddin, A., Murthy, J. K., Hamid, A., & Ugusman, A. (2022). Effects of *Piper sarmentosum* Roxb. on hypertension and diabetes

- mellitus: A systematic review and meta-analysis. *Frontiers in Pharmacology*, 13, 976247. <https://doi.org/10.3389/fphar.2022.976247>
7. Purba, R. A. P., Paengkoum, S., & Paengkoum, P. (2021). Development of a simple high-performance liquid chromatography-based method to quantify synergistic compounds and their composition in dried leaf extracts of *Piper sarmentosum* Roxb. *Separations*, 8, 152. <https://doi.org/10.3390/separations8090152>
 8. Ridditid, W., Rattanaprom, W., Thaina, P., Chittrakarn, S., & Sunbhanich, M. (1998). Neuromuscular blocking activity of methanolic extract of *Piper sarmentosum* leaves in the rat phrenic nerve-hemidiaphragm preparation. *Journal of Ethnopharmacology*, 61, 135–142. [https://doi.org/10.1016/S0378-8741\(98\)00025-7](https://doi.org/10.1016/S0378-8741(98)00025-7)
 9. Scott, D. L., Wolfe, F., & Huizinga, T. W. (2010). Rheumatoid arthritis. *The Lancet*, 376, 1094–1108. [https://doi.org/10.1016/S0140-6736\(10\)60826-4](https://doi.org/10.1016/S0140-6736(10)60826-4)
 10. Sim, K.-M., Mak, C.-N., & Ho, L.-P. (2009). A new amide alkaloid from the leaves of *Piper sarmentosum*. *Journal of Asian Natural Products Research*, 11, 757–760. <https://doi.org/10.1080/10286020903058933>
 11. Soeken, K. L., Miller, S. A., & Ernst, E. (2003). Herbal medicines for the treatment of rheumatoid arthritis: A systematic review. *Rheumatology*, 42, 652–659. <https://doi.org/10.1093/rheumatology/keg183>
 12. Srirangan, S., & Choy, E. H. (2010). The role of interleukin 6 in the pathophysiology of rheumatoid arthritis. *Therapeutic Advances in Musculoskeletal Disease*, 2, 247–256. <https://doi.org/10.1177/1759720X10378372>
 13. Suhana Mohd Ramli, E., Nirwana Soelaiman, I., Othman, F., Ahmad, F., Nazrun Shuib, A., Mohamed, N., et al. (2012). The effects of *Piper sarmentosum* water extract on the expression and activity of 11 β -hydroxysteroid dehydrogenase type 1 in bones with excessive glucocorticoids. *Iranian Journal of Medical Sciences*, 37, 39–46.
 14. Sun, X., Chen, W., Dai, W., Xin, H., Rahmand, K., Wang, Y., et al. (2020). *Piper sarmentosum* Roxb.: A review on its botany, traditional uses, phytochemistry, and pharmacological activities. *Journal of Ethnopharmacology*, 263, 112897. <https://doi.org/10.1016/j.jep.2020.112897>
 15. Takeuchi, T., Yoshida, H., & Tanaka, S. (2021). Role of interleukin-6 in bone destruction and bone repair in rheumatoid arthritis. *Autoimmunity Reviews*, 20, 102884. <https://doi.org/10.1016/j.autrev.2021.102884>
 16. Udenze, E. C. C., Ezirim, A., Ihedimbu, C. P., & IHEME, C. (2014). Effects of oral administration of *Garcinia kola* seeds on haematological and defence parameters of diabetic rats. *American Journal of Biochemistry and Molecular Biology*, 4, 167–175. <https://doi.org/10.3923/ajbmb.2014.167.175>
 17. Ugusman, A., Zakaria, Z., Hui, C. K., & Nordin, N. A. M. M. (2011). *Piper sarmentosum* inhibits ICAM-1 and Nox4 gene expression in oxidative stress-induced human umbilical vein endothelial cells. *BMC Complementary and Alternative Medicine*, 11, 31. <https://doi.org/10.1186/1472-6882-11-31>

18. Ugusman, A., Zakaria, Z., Hui, C. K., Nordin, N. A. M. M., & Mahdy, Z. A. (2012). Flavonoids of *Piper sarmentosum* and its cytoprotective effects against oxidative stress. *EXCLI Journal*, 11, 705–714.
19. Umar, H. I., Josiah, S. S., Saliu, T. P., Jimoh, T. O., Ajayi, A., & Danjuma, J. B. (2021). In silico analysis of the inhibition of the SARS-CoV-2 main protease by some active compounds from selected African plants. *Journal of Taibah University Medical Sciences*, 16, 162–176. <https://doi.org/10.1016/j.jtumed.2020.12.005>
20. Vimala, S., Mohd, I. A., Abdull, R. A., & Rohana, S. (2003). Natural antioxidants: *Piper sarmentosum* (kadok) and *Morinda elliptica* (mengkudu). *Malaysian Journal of Nutrition*, 9, 41–51.
21. Wang, D. F., Zhou, L. L., Zhou, H. L., Hou, G. Y., Zhou, X., & Li, W. (2017). Effects of *Piper sarmentosum* extract on growth performance, antioxidant capability and immune response in weaned piglets. *Journal of Animal Physiology and Animal Nutrition*, 101, 105–112. <https://doi.org/10.1111/jpn.12517>
22. Ware, I., Franke, K., Dube, M., Ali El Enshasy, H., & Wessjohann, L. A. (2023). Characterization and bioactive potential of secondary metabolites isolated from *Piper sarmentosum* Roxb. *International Journal of Molecular Sciences*, 24, 1328. <https://doi.org/10.3390/ijms24021328>
23. Xu, X., Xu, H., Shang, Y., Zhu, R., Hong, X., Song, Z., et al. (2021). Development of the general chapters of the Chinese Pharmacopoeia 2020 edition: A review. *Journal of Pharmaceutical Analysis*, 11, 398–404. <https://doi.org/10.1016/j.jpha.2021.05.001>
24. Yeo, E. T. Y., Wong, K. W. L., See, M. L., Wong, K. Y., Gan, S. Y., & Chan, E. W. L. (2018). *Piper sarmentosum* Roxb. confers neuroprotection on beta-amyloid (A β)-induced microglia-mediated neuroinflammation and attenuates tau hyperphosphorylation in SH-SY5Y cells. *Journal of Ethnopharmacology*, 217, 187–194. <https://doi.org/10.1016/j.jep.2018.02.025>
25. Yeoh, H.-H., & Wong, P.-F. M. (1993). Food value of lesser utilised tropical plants. *Food Chemistry*, 46, 239–241. [https://doi.org/10.1016/0308-8146\(93\)90113-T](https://doi.org/10.1016/0308-8146(93)90113-T)
26. Yoshida, Y., & Tanaka, T. (2014). Interleukin-6 and rheumatoid arthritis. *BioMed Research International*, 2014, 698313. <https://doi.org/10.1155/2014/698313>
27. Zainab, B., Ayaz, Z., Alwahibi, M. S., Khan, S., Rizwana, H., Soliman, D. W., et al. (2020). In silico elucidation of *Moringa oleifera* phytochemicals against diabetes mellitus. *Saudi Journal of Biological Sciences*, 27, 2299–2307. <https://doi.org/10.1016/j.sjbs.2020.04.002>
28. Zainal Ariffin, S. H., Wan Omar, W. H. H., Zainal Ariffin, Z., Safian, M. F., Senafi, S., & Megat Abdul Wahab, R. (2009). Intrinsic anticarcinogenic effects of *Piper sarmentosum* ethanolic extract on a human hepatoma cell line. *Cancer Cell International*, 9, 6. <https://doi.org/10.1186/1475-2867-9-6>
29. Zainol Abidin, I. Z., Johari, A. N., Yazid, M. D., Zainal Ariffin, Z., Eziwar Dyari, H. R., & Zainal Ariffin, S. H. (2023). Osteogenic potential and bioactive profiles of *Piper*

- sarmentosum* ethanolic extract-treated stem cells. *Pharmaceuticals*, 16, 708. <https://doi.org/10.3390/ph16050708>
30. Zakaria, Z. A., Patahuddin, H., Mohamad, A. S., Israf, D. A., & Sulaiman, M. R. (2010). In vivo anti-nociceptive and anti-inflammatory activities of the aqueous extract of the leaves of *Piper sarmentosum*. *Journal of Ethnopharmacology*, 128, 42–48. <https://doi.org/10.1016/j.jep.2009.12.021>
 31. Zeng, L., Yang, T., Yang, K., Yu, G., Li, J., Xiang, W., et al. (2022). Efficacy and safety of curcumin and *Curcuma longa* extract in the treatment of arthritis: A systematic review and meta-analysis of randomized controlled trials. *Frontiers in Immunology*, 13, 891822. <https://doi.org/10.3389/fimmu.2022.891822>
 32. Zhang, Y., Mao, X., Li, W., Chen, W., Wang, X., Ma, Z., et al. (2021). *Tripterygium wilfordii*: An inspiring resource for rheumatoid arthritis treatment. *Medicinal Research Reviews*, 41, 1337–1374. <https://doi.org/10.1002/med.21762>
 33. Abdelmohsen, U. R., Bayer, K., & Hentschel, U. (2014). Diversity, abundance and natural products of marine sponge-associated actinomycetes. *Natural Product Reports*, 31, 381–399. <https://doi.org/10.1039/c3np70111e>
 34. Abdollahpour-Alitappeh, M., Lotfinia, M., Gharibi, T., Mardaneh, J., Farhadihosseinabadi, B., Larki, P., et al. (2019). Antibody-drug conjugates (ADCs) for cancer therapy: Strategies, challenges, and successes. *Journal of Cellular Physiology*, 234, 5628–5642. <https://doi.org/10.1002/jcp.27419>
 35. AlAjmi, M. F., Rehman, M. T., Hussain, A., & Rather, G. M. (2018). Pharmacoinformatics approach for the identification of polo-like kinase-1 inhibitors from natural sources as anti-cancer agents. *International Journal of Biological Macromolecules*, 116, 173–181. <https://doi.org/10.1016/j.ijbiomac.2018.05.023>
 36. Aljofan, M., & Riethmacher, D. (2019). Anticancer activity of metformin: A systematic review of the literature. *Future Science OA*, 5, FSO410. <https://doi.org/10.2144/fsoa-2019-0053>
 37. Amaral, M. E. A., Nery, L. R., Leite, C. E., de Azevedo Junior, W. F., & Campos, M. M. (2018). Pre-clinical effects of metformin and aspirin on the cell lines of different breast cancer subtypes. *Investigational New Drugs*, 36, 782–796. <https://doi.org/10.1007/s10637-018-0568-y>
 38. Arba, M., Yamin, Ihsan, S., & Tjahjono, D. H. (2018). Computational approach toward targeting the interaction of porphyrin derivatives with Bcl-2. *Journal of Applied Pharmaceutical Science*, 8(12), 060–066.
 39. Bachmann, B. O., Van Lanen, S. G., & Baltz, R. H. (2014). Microbial genome mining for accelerated natural products discovery: Is a renaissance in the making? *Journal of Industrial Microbiology & Biotechnology*, 41, 175–184. <https://doi.org/10.1007/s10295-013-1389-9>

40. Barbuti, A., & Chen, Z.-S. (2015). Paclitaxel through the ages of anticancer therapy: Exploring its role in chemoresistance and radiation therapy. *Cancers*, 7, 2360–2371. <https://doi.org/10.3390/cancers7040897>
41. Beck, A., Goetsch, L., Dumontet, C., & Corvaia, N. (2017). Strategies and challenges for the next generation of antibody-drug conjugates. *Nature Reviews Drug Discovery*, 16, 315–337. <https://doi.org/10.1038/nrd.2016.268>
42. Belwal, T., Bisht, A., Devkota, H. P., Ullah, H., Khan, H., Pandey, A., et al. (2020). Phytopharmacology and clinical updates of *Berberis* species against diabetes and other metabolic diseases. *Frontiers in Pharmacology*, 11, 41. <https://doi.org/10.3389/fphar.2020.00041>
43. Bisen, P. S. (2016). Experimental and computational approaches in leveraging natural compounds for network-based anti-cancer medicine. *Journal of Cancer Science Research*, 2, e103. <https://doi.org/10.4172/2576-1447.1000e103>
44. Boucher, I. W., McMillan, P. J., Gabrielsen, M., Akerman, S. E., Brannigan, J. A., Schnick, C., et al. (2006). Structural and biochemical characterization of a mitochondrial peroxiredoxin from *Plasmodium falciparum*. *Molecular Microbiology*, 61, 948–959. <https://doi.org/10.1111/j.1365-2958.2006.05303.x>
45. Cerella, C., Muller, F., Gaigneaux, A., Radogna, F., Viry, E., Chateauvieux, S., et al. (2015). Early downregulation of Mcl-1 regulates apoptosis triggered by cardiac glycoside UNBS1450. *Cell Death & Disease*, 6, e1782. <https://doi.org/10.1038/cddis.2015.134>
46. Chakraborti, C. (2011). Vitamin D as a promising anticancer agent. *Indian Journal of Pharmacology*, 43, 113–120. <https://doi.org/10.4103/0253-7613.77335>
47. Chandra, S. (2012). Endophytic fungi: Novel sources of anticancer lead molecules. *Applied Microbiology and Biotechnology*, 95, 47–59. <https://doi.org/10.1007/s00253-012-4128-7>
48. Charlop-Powers, Z., Milshteyn, A., & Brady, S. F. (2014). Metagenomic small molecule discovery methods. *Current Opinion in Microbiology*, 19, 70–75. <https://doi.org/10.1016/j.mib.2014.05.021>
49. Charmsaz, S., Prencipe, M., Kiely, M., Pidgeon, G., & Collins, D. (2018). Innovative technologies changing cancer treatment. *Cancers*, 10, 208. <https://doi.org/10.3390/cancers10060208>
50. Chavda, V. P., Mohapatra, S. S., Ranjan, S., Dasgupta, N., Mishra, R. K., and Thomas, D. S. (2019). “Chapter 4 - Nanobased Nano Drug Delivery: A Comprehensive Review,” in *Micro And Nano Technologies*. Elsevier, 69–92. doi:10.1016/B978-0-12-814029-1.00004-1